pda.org

Sterilization Processes – 2024 What do we do now?

James Agalloco

President

Agalloco & Associates Inc.



PDA Aseptic Processing of Biopharmaceuticals Conference 2024





EMA / Annex 1 – Sterilisation FDA / USP – Sterilization





COPYRIGHT © PDA 2024

pda.org



A Growing and Unfortunate Divide

- FDA guidelines are flexible with exceptions rather common.
- Multiple layers of internal review & field inspection.
- USP extensive sterilization content (50+ pages) that is relatively current (2012-2019). Includes all major sterilization methods.
- Many guidelines with general requirements with few specifics.
- Interpretive What to do!

- EMA guidelines are more restrictive with fewer exceptions.
- Reviewer and filed inspector more often the same person.
- EP limited sterilization content (4-5 pages) from 2008. Limited to steam, dry heat, radiation and ETO.
- Annex 1 includes many specific requirements restricting flexibility of execution.
- Substantive How to do!







USP Sterilization <1229. -01 - 12

- 1229.00 Introduction & General Principles
- 1229.01 Steam Sterilization by Direct Contact
- 1229.02 Moist Heat Sterilization of Aqueous Liquids
- 1229.03 Monitoring of Bioburden
- 1229.04 Sterilizing Filtration of Liquids
- 1229.05 Biological Indicators for Sterilization
- 1229.06 Liquid Phase Sterilization
- 1229.07 Gaseous Sterilization
- 1229.08 Dry Heat Sterilization
- 1229.09 Physicochemical Integrators and Indicators for Sterilization
- 1229.10 Radiation Sterilization
- 1229.11 Vapor Phase Sterilization
- 1229.12 New Sterilization Methods





pda.org



USP Sterilization <1229. -13 - 20

- 1229.13 Sterilization-in-Place
- 1229.14 Sterilization Cycle Development
- 1229.15 Sterilizing Filtration of Gases
- 1229.16 Prion Inactivation
- 1229.17 Mycoplasma Sterilization
- 1229.18 Viral Clearance
- 1229.19 Simultaneous Decontamination & Sterilization
- 1229.20 Sterilization: Pre-, Mid-, and Post-Process Considerations

Related Chapters

- 55 Biological Indicators Resistance Performance Tests Revised
- 56 Methods for Determination of Resistance of Microorganisms to Sterilization Processes all new content
- 1211 Sterilization & Sterility Assurance all sterilization process related content deleted, substantial number of other changes



Annex 1 - Sterilization / Sterilisation

- The content in this section is heavily influenced by its Orange Guide origins; HTM10, EN285 and pretty much anything written on the subject in the UK!
- US sterilization practices differ markedly. Which practices will your investigator / auditor / reviewer accept? Harmonization on which set of expectations?
- Understand that the *European Pharmacopiea* includes only steam, dry heat, gas (ETO) or gamma irradiation.
- Compare this to USP which includes moist heat, dry heat, gas (ETO, O₃, ClO₂, NO₂), liquids, radiation (gamma, X-ray, E-beam), and vapors (H₂O₂, peracetic acid).





Sterilization / Decontamination with H₂O₂

- In 2018, an MHRA blog post stated, "... our current stance is that VHP cannot be used to sterilise critical items."
- The concern was raised regarding the reports of 'rogue' BI results suggesting the vapor process is inherently fragile.
- This has curtailed H_2O_2 use in various applications with isolator decontamination / sterilization the most impacted.
- The problem may be due to false beliefs, inadequate system design, BI configuration and more.
- FDA has approved deep vacuum H₂O₂ sterilization processes which are not the same.





Sterilization of Isolator Installed Equipment

- *"Particular attention should be given when the adopted product sterilisation method is not described in the current edition of the Pharmacopoeia, ..."* [8.37]
- Vapor sterilization for RABS & isolators where "e.g. sterilised items such as stopper bowls and guides, and sterilised components" are sterilized would be non-compliant. [5.5]
- This is a major challenge which cannot be easily addressed. It increases manual activity and contamination risk as well.





Vapor Sterilization / Decontamination Practices Troubles and Traumas





COPYRIGHT © PDA 2024



Process Difficulties due to 'Rogue' BI's

- Positive BI results in sterilization / decontamination systems have caused difficulties in initial / periodic validation studies.
- Project delays and supply interruptions have resulted.
- The MHRA blog 2018 suggested imposing a ban on VPHP for the sterilization of product contact surfaces. EMA Annex 1 2024 is aligned with that belief. FDA is characteristically silent.
- Unknown variations in BI's are suspected to be 'root cause'.
- Do 'rogues' exist and if so, can they be eliminated?







Vapor Processes & Thermodynamics

- All sterilization process seek uniform conditions throughout.
- Vapor processes also tend towards equilibrium, but this leads to condensation and the inevitable presence of two phases.
- This results in non-uniformity in phase and concentration and inconsistent lethality across the system.
- Aggressive mixing could reduce variation, but current designs are rarely sufficient.
- Most two-phase vapor systems have insufficient mixing, and some rely totally on unidirectional air, which is the exact opposite of what is needed to create uniformity.



Gases, Liquids & Vapors





Physical-Chemical Reality of VPHP Processes

- The process may not reach equilibrium due to continued heat input during process and temperature variations (inlet to exhaust) across system.
- Temperature varies with time and location, and so will phase and concentrations. The amount of condensed H_2O_2 and H_2O varies with local temperature.
- Precise determination of conditions across the entire system and its surfaces is impossible.











Dry – avoid condensation

- Vapors are gases, it's always a single-phase
- Gas H_2O_2 kills quicker than liquid H_2O_2
- Avoid condensation
- Mixing is not a concern
- Less H₂O₂ used

Wet – encourage condensation

- Vapors are 2-phase mixtures of gases and liquids
- Liquid H₂O₂ kills quicker than gas H₂O₂
- Condensation is desirable
- Mixing for greater uniformity
- More H₂O₂ used

It doesn't matter what we try to do, both phases will always be present

'Dry' vs 'Wet'













Another Way to Look at Vapor Processes

- It's understood that process conditions directly impact process lethality.
- For example, in thermal sterilization a higher process temperature results in a lower D-value or a steeper death curve. This is true for all processes, the more aggressive the conditions the easier it is to kill microorganisms.
- If the conditions are non-uniform, then there's the potential for different lethal effects in different places.
- Let's apply that thinking to H₂O₂ vapor processes.





Time



What's the Real Root Cause of Positive Results?It's a 'Rogue'It's a Non-Uniform Process



The similarity in appearance it not a coincidence, it serves to inform that there's no reason to believe that 'rogue' BIs are the only cause of anomalous results.



The Unfortunate Reality

- BI configurations are often sub-optimal.
- The unavoidable variations in vapor H₂O₂ processes may cause anomalous results unrelated to the biological indicator but due to varying conditions across the system.
- The complexities of the vapor process can give the appearance of 'rogues' even where every BI is identical and free of defects.
- The kill rate curves shown are simplified. There's an infinity of them between the extremes and kill rates at any location will change during the process.
- The death rate curves are not straight lines.







Use the most suitable Biological Indicators

- Pick a reliable supplier with sufficient experience.
- Avoid use of planar stainless-steel coupons, use non-cellulose fiber strips as the BI substrate.
- Use a non-Tyvek protective wrap or remove the wrap before exposure (false positive results with *G. stearothermophilus* spores are nearly impossible).
- Don't bother with multiple BI's in locations, its preferable to use the same total number of BI's in more locations.
- Use a challenge level of 10⁴ spores/strip unless sterilization is required, use 10⁶ spores/strip for those items and surfaces.





Use the most lethal process you can

- Forget the idea that condensate is to be avoided, that's likely wrong. Use enough H_2O_2 ; but not so much as to have drops form on surfaces.
- Dehumidification which removes H_2O is not beneficial, it will not alter the amount of H_2O_2 that will condense.
- Use more internal mixing, unidirectional air works against uniformity and should not be used. This works to advantage in aeration as well. The more mixing the better.
- Aeration times may not change much as micro-condensation has always been present, just not always enough to effect consistent kill at all locations.





Annex 1 / FDA Terminal Sterilization





pda.org

Annex 1 Terminal Sterilization

- The Annex 1 included content is non-controversial.
- The elephant in the room is EMA's Guideline on the sterilisation of the medicinal product, active substance, excipient and final container. EMA\CHMP\CVMP\QWP\850374\2015





Sterilization – USP <1229>

- "Articles intended to be sterile must attain a ≤10⁻⁶ probability of a nonsterile unit, i.e., less than than or equal to 1 chance in 1 million that viable bioburden microorganisms are present."
- What does this mean in the real world?
 - The process needs to reliably destroy the bioburden.
 - No minimum F₀ requirement.
 - No minimum time or temperature.
 - No specified BI population or kill.
 - No defined sterilization process.
- It allows for flexibility in sterilization process design to achieve the required result free of arbitrary requirements.





Calculation of PNSU / SAL



where:

- Nu PNSU / SAL =
- D D-value of the natural bioburden =
- F = F-value (lethality) of the process
- No bioburden population =







The Terminal Sterilization Dilemma

• The goal of sterilization must be balanced against the need to maintain the product's essential quality attributes.







Microorganisms and Growth



Microorganism	D ₆₀ – Pflug reported (minutes)	z-value Pflug reported (°C)	D ₈₀ – estimated** (minutes)	D ₁₀₀ – estimated** (minutes)	Assumed No**** (CFU)	Estimated PNSU @ 80°/20 minutes***	Estimated PNSU @ 100°C / 5 minutes***
Aeromonas hydrophilia	0.066	6.0	0.00007	0.0000007	100	10 ^{-285,000}	10 ^{-71,400,000}
Campylobacter jejuni	0.35	6.5	0.0004	0.0000004	100	10 ^{-50,000}	10 ^{-12,500,000}
Escherichia coli	2.0	5.0	0.0002	0.00000002	100	10 ^{-100,000}	10 ^{-250,000,000}
Listeria monocytogenes	16.7	6.5	0.02	0.00002	100	10 ^{-1,000}	10 ^{-50,000}
Salmonella species	2.5	5.6	0.003	0.000003	100	10 ^{-6,600}	10 ^{-1,666,000}
Staphylococcus aureus	3.0	9.5	0.03	0.0003	100	10 ⁻⁶⁰⁰	10 ^{-16,600}
Vibrio species	2.6	6.0	0.003	0.000003	100	10 ^{-6,600}	10 ^{-1,666,000}
Yersinia enterocolitica	5.0	5.0	0.0005	0.00000005	100	10 ^{-40,000}	10 ^{-100,000,000}

Table 1 – Calculated PNSU for selected pathogens	at 80°C / 20 min & 100°C for 5 minutes*
--	---

D₆₀ and z-value data in this table from Pflug.¹⁸ *

 $\mathsf{D}_{so},$ and $\mathsf{D}_{100}\mathsf{values}$ calculated from Pflug and rounded off for ease of calculation. **

*** PNSU values rounded off for ease of comprehension

Note that the No values have no meaningful impact on the estimated PNSU. ****



Some Successful Terminal Sterilization Projects





Thanks for Your Attention!



Questions?

Contact Information

James Agalloco Agalloco & Associates PO Box 899 Belle Mead, NJ 08502

jagalloco@aol.com



pda.org